

Suppl Table S1. Major QTLs identified in MT-RILs, backcrossed lines, and the MPH datasets

Trait	Name	Chr.	Interval	RILs				BCF _s				MPH				QTL type		
				LOD	a	R2(%)	PVE (%)	LOD	a+d	R2(%)	PVE (%)	LOD	d	R2(%)	PVE (%)			
CL	<i>qCL1.1</i>	1	RM600-S01054	3.05	2.87	2.4	89.2										A	
	<i>qCL1.2</i>	1	S01143A-S01147														OD	
	<i>qCL1.3</i>	1	S01147-S01157B	50.18	-14.14	57.2		44.18	9.56	65.7			4.97	2.37	10.5		A	
	<i>qCL3.1</i>	3	RM7-RM251	6.27	4.40	5.5		5.78	-3.01	6.5							A	
	<i>qCL6.1</i>	6	S06087-S06100	9.92	5.01	7.2		6.07	-2.68	5.2							A	
	<i>qCL7.1</i>	7	S07035-S07048	8.98	5.35	8.2											A	
	<i>qCL7.2</i>	7	S07048-S07050A										3.76	1.99	7.3		OD	
	<i>qCL8.1</i>	8	S08059-S08066	3.22	2.58	1.9											A	
	<i>qCL10.1</i>	10	S10026C-RM184	5.2	-4.12	4.9											A	
	<i>qCL11.1</i>	11	RM21-RM206	3.74	3.08	2.7											A	
	DTH	<i>qDTH1.1</i>	1	RM1-S01027					73.7	2.91	0.83	2.4	38.7				no analysis	A
<i>qDTH3.1</i>		3	S03137-S03145	7.4	2.35	5.7											A	
<i>qDTH6.1</i>		6	RM527-RM3183					27.68	3.24	36.3							A	
<i>qDTH7.2</i>		7	S07050A-S07055B	43.29	7.68	61.1											A	
<i>qDTH11.1</i>		11	RM206-RM187	7.66	2.58	6.9											A	
PN	<i>qPN3.1</i>	3	S03033-S03024A	3.22	0.64	6.8	14.0						14.0			7.5	A	
	<i>qPN3.2</i>	3	S03041-RM7					2.57	-0.30	5.7							A	
	<i>qPN5.1</i>	5	RM146-RM430					4.33	0.37	8.4							A	
	<i>qPN5.2</i>	5	RM163-S05087										3.27	0.38	7.5		OD	
	<i>qPN6.1</i>	6	RM5087-S06084	3.26	0.66	7.2											A	
PL	<i>qPL1.1</i>	1	S01143A-S01147	6.84	-0.98	10.2	40.2						43.1			24.5	A	
	<i>qPL1.2</i>	1	S01147-S01157B					6.44	0.56	11.2							A	
	<i>qPL4.1</i>	4	RM348-S04120					13.54	0.86	26.7							A	
	<i>qPL4.2</i>	4	S04120-S04128										7.58	0.57	11.7		OD	
	<i>qPL5.1</i>	5	S05062-RM146	5.49	0.94	9.5											A	
	<i>qPL6.1</i>	6	S06087-S06100	3.85	0.69	5.0		2.72	-0.38	5.2							A	
	<i>qPL7.1</i>	7	S07035-S07048	8.41	1.20	15.5							7.68	0.59	12.8		D	
	<i>qSN1.1</i>	1	S01022-RM1	3.87	11.87	5.0	64.0						40.6	15.76	15.96	26.7	32.1	OD
SN	<i>qSN1.2</i>	1	RM428-RM259					10.5	-13.53	13.2							A	
	<i>qSN1.3</i>	1	RM259-RM600	5.76	18.71	12.5											A	
	<i>qSN1.4</i>	1	S01143A-S01147	4.89	-12.63	5.7											A	
	<i>qSN2.1</i>	2	RM438-RM424	2.52	10.44	3.9											A	
	<i>qSN2.2</i>	2	RM492-RM71					6.83	-9.75	6.8							A	
	<i>qSN3.1</i>	3	S03010B-RM231					3.14	6.90	3.4							A	
	<i>qSN4.1</i>	4	RM303-S04087A					4.7	9.10	6.0							A	
	<i>qSN4.2</i>	4	RM348-S04120	5.84	-14.42	7.4											A	
	<i>qSN6.1</i>	6	RM50-RM527					9.4	12.49	11.2							A	
	<i>qSN6.2</i>	6	S06087-S06100	3.28	10.93	4.3											A	
	<i>qSN7.1</i>	7	S07048-S07050A	13.99	24.87	22.1											A	
	<i>qSN7.2</i>	7	S07080-RM234	2.79	-10.91	4.3											A	
	<i>qSN8.1</i>	8	RM447-S08112										5.04	-7.21	5.4		UD	
	SF	<i>qSF1.1</i>	1	S01147-S01157B	9.42	-5.44	18.4	28.2						48.2			no analysis	A
		<i>qSF6.1</i>	6	S06015-S06018					11.63	-9.69	23.9							A
		<i>qSF6.2</i>	6	RM50-RM527					5.34	-6.47	10.6							A
<i>qSF9.1</i>		9	S09055-S09058	3.64	3.96	9.8											A	
TGW	<i>qTGW2.1</i>	2	S02010-S02020					32.3	5.93	0.35	5.9	17.2				20.2	A	
	<i>qTGW2.2</i>	2	S02043-RM438	11.41	-0.99	17.0											A	
	<i>qTGW3.1</i>	3	S03145-S03152B										2.94	0.37	9.1		OD	
	<i>qTGW8.1</i>	8	S08020A-RM25	3.71	-0.56	5.3											A	
	<i>qTGW9.1</i>	9	RM108-RM205										3.68	-0.41	11.1		UD	
<i>qTGW12.1</i>	12	S12109A-RM17	6.35	-0.76	10.0		9.56	0.49	11.3							A		
GYP	<i>qGYP1.1</i>	1	RM428-RM259	6.18	2.31	8.4	54.9						25.1			39.3	A	
	<i>qGYP1.2</i>	1	RM259-RM600										3.85	1.77	5.6		OD	
	<i>qGYP1.3</i>	1	S01147-S01157B	21.83	-4.60	33.1											A	
	<i>qGYP4.1</i>	4	RM348-S04120					3.96	2.00	8.1							A	
	<i>qGYP5.1</i>	5	S05095-RM480										6.95	2.43	10.6		OD	
	<i>qGYP6.1</i>	6	S06015-S06018					6.66	-2.91	17.0			8.24	-2.86	14.6		UD	
	<i>qGYP7.1</i>	7	S07035-S07048	8.96	3.01	14.1											A	
	<i>qGYP8.1</i>	8	S08059-S08066										2.29	1.26	2.8		OD	
	<i>qGYP10.1</i>	10	S10019-S10026C										3.77	-1.80	5.8		UD	
	PYP	<i>qPYP1.1</i>	1	S01143A-S01147	9.09	-2.70	11.5	38.7						32.5			35.4	A
<i>qPYP1.2</i>		1	S01160-RM104					5.11	2.17	6.2							A	
<i>qPYP6.1</i>		6	RM50-RM527					11.25	4.46	26.2			12.83	4.55	31.7		OD	
<i>qPYP7.1</i>		7	S07035-S07048	16.05	4.14	27.2											A	
<i>qPYP11.1</i>		11	S11112-S11117B										2.6	1.55	3.7		OD	

Suppl Table S2. EpQTLs of MT-RILs and BCF₁ populations and MPH datasets of BCF₁s.

Trait	Chr.	Interval i	Chr.	Interval j	LOD	Ai	Aj	AAij	H ² (Ai)	H ² (Aj)	H ² (AAij)	PVE (%)
RIL	CL	RM271-RM258		RM167-RM202	4.76	-2.47*		4.14	0.0143	0.0003	0.0402	4.02
	DTH	RM104-RM414		RM418-RM11	4.82			-2.13	0.0002	0	0.0138	2.12
		RM527-RM3183		S07050A-S07055B	44.11		7.77***	-1.56	0.0012	0.1834	0.0074	
	PN	S03027-S03041		RM163-S05087	6.26			0.75	0.0183	0.0043	0.0946	29.96
		RM347-S03136		RM5087-S06084	5		0.44**	-0.68	0.0132	0.0325	0.0777	
		S07048-S07050A		RM429-S07118	6.01			0.88	0.0061	0.0043	0.1273	
	SF	S01022-RM1		S08059-S08066	4.89		2.33*	3.31	0.0104	0.0319	0.0644	17.67
		RM1-S01027		S11004A-RM332	5.44			-4.37	0.0011	0.0043	0.1123	
	TGW	RM259-RM600		RM447-S08112	5.48			-0.62	0.0004	0.005	0.0669	18.53
		S02010-S02020		RM289-S05062	5.5	-0.4**		0.42	0.0278	0.007	0.0307	
		S03137-S03145		RM206-RM187	5.9			-0.71	0.0002	0	0.0877	
	GYP	S02000A-RM485		S12056-RM260	5.5			-2.36	0.0004	0.0014	0.0738	13.73
PYP	RM346-RM336		S11059-S11070	5.42		1.15*	-2.19	0.0001	0.0175	0.0635		
	S01158-S01160		S12109B-S12094	4.71		-1.48**	1.7	0.0023	0.037	0.0488	20.08	
	RM11-S07071		S11059-S11070	9.67			-3	0.0024	0.0017	0.152		
F1	CL	S01147-S01157B		S07035-S07048	48.7	*-9.85***		-1.92	0.7711	0.0001	0.0293	2.93
	DTH	S09045-S09049		RM167-RM202	5.67			-1.3	0.0042	0.006	0.0781	7.81
	PN	RM428-RM259		RM4A-S12011B	5.85			-0.46	0.0014	0.0109	0.1182	32.92
		S03137-S03145		S08038-RM72	4.7			0.33	0.0014	0.0109	0.0608	
		S06019B-S06031		RM248-RM420	5.03	0.24**	-0.2*	-0.33	0.0322	0.0223	0.0608	
		S10019-S10026C		RM167-RM202	6.28			0.4	0.0068	0.002	0.0894	
	PL	RM1-S01027		S02000A-RM485	4.88			-0.52	0.0005	0.003	0.0823	12.4
		RM527-RM3183		RM270-RM235	5.5	-0.24*		-0.37	0.0175	0	0.0417	
	SN	S02085-S02088		S05000-RM267	4.75			7.3	0.0011	0.0022	0.0385	10.1
		S03010B-RM231		S10072-RM228	8.95	-7.16***	-4.23**	-6.71	0.0371	0.0129	0.0326	
		RM303-S04087A		RM31-RM334	6.15	-7.74***		-6.43	0.0433	0.0001	0.0299	
	SF	RM212-S01140		RM201-RM108	5.37			-4.86	0.0046	0.0004	0.0482	14.53
		S03033-S03024A		S11112-S11117B	4.69			3.37	0.0001	0.0036	0.0232	
		RM234-RM118		RM270-RM235	5.17			4.04	0.0023	0.0008	0.0333	
		S07101-S07103		S09065-S09075B	5.04			-4.46	0.0004	0.0057	0.0406	
	TGW	RM263-RM250		S03145-S03152B	5.74			0.39	0.0019	0.0137	0.0811	15.81
		S03145-S03152B		S11106-S11112	5.81			-0.38	0.0065	0	0.077	
	PYP	RM341-S02081B		RM480-S05116	4.86			-2.05	0.006	0	0.0476	8.55
	S03046-S03055		S06019B-S06031	4.95			-1.83	0.0001	0.0076	0.0379		
MPH	CL	S01158-S01160		S09049-S09055	5.44	-1.4*		-1.86	0.0406	0.0123	0.0717	7.17
	DTH	RM246-S01127		S09065-S09075B	5.74			-1.44	0.0003	0.0001	0.0477	13.43
		RM204-S06015		S08000-RM337	7.48			-1.67	0	0.0033	0.0641	
		RM527-RM3183		S07048-S07050A	33.33	-2.53***	-3.02***	0.99	0.1471	0.2096	0.0225	
	PL	RM492-RM71		S12029-RM277	5.36			0.57	0.0033	0.0131	0.1061	17.11
		S05030B-S05045		S05062-RM146	5.38			0.49	0.0005	0.0026	0.065	
	SF	RM1-S01027		S03164-RM85	6.18	3.18**		4.39	0.0167	0.0008	0.0318	14.62
		RM488-RM246		S12056-RM260	5.66			-4.12	0.0011	0.0012	0.028	
		S02085-S02088		RM481-S07024	7.41			-5.28	0.0017	0.0005	0.046	
		S03033-S03024A		S11112-S11117B	5.43			3.34	0.0004	0.0081	0.0184	
		RM489-S03020		RM50-RM527	17.78		10.25***	3.65	0.0025	0.1732	0.022	
	TGW	RM251-S03046		S11106-S11112	5.87			0.35	0.0111	0.0151	0.0944	19.43
		RM11-S07071		S11077-RM21	4.95			0.36	0	0.0062	0.0999	
	GYP	RM485-RM211		RM251-S03046	5.15			-2.02	0.0008	0.0006	0.0512	14.04
		S02010-S02020		S03136-S03137	5.58			2.02	0.0006	0	0.0512	
	S05087-S05095		S09034-RM105	6.89	-2.45***		1.74	0.0753	0.0029	0.038		

Suppl Table S3. Comparison of the heterosis QTLs in two indica-japonica RILs

Trait	DT						MT					
	QTL ^a	Chr.	Flanking markers	LOD ^b	a ^c	R ² (%) ^d	QTL ^a	Chr.	Flanking markers	LOD ^b	a ^c	R ² (%) ^d
CL							<i>qCL1.2</i>	1	S01143A-S01147	4.97	2.37	10.5
							<i>qCL7.2</i>	7	S07048-S07050A	3.76	1.99	7.3
	<i>qCL9.1</i>	9	S09000A-S09006	4.71	2.11	15.1						
PN							<i>qPN5.2</i>	5	RM163-S05087	3.27	0.38	7.5
PL	<i>qPL4.1</i>	4	S04077A-RM451	5.38	0.45	9.4						
							<i>qPL4.2</i>	4	S04120-S04128	7.58	0.57	11.7
							<i>qPL7.1</i>	7	S07035-S07048	7.68	0.59	12.8
	<i>qPL8.1</i>	8	RM25-RM72	7.83	0.58	15.4						
SN							<i>qSN1.1</i>	1	S01022-RM1	15.76	15.96	26.7
							<i>qSN8.1</i>	8	RM447-S08112	5.04	-7.21	5.4
	<i>qSN9.1</i>	9	S09000A-S09006	2.56	6.38	6.2						
TGW	<i>qTGW1.1</i>	1	RM297-S01140	6.12	0.53	13.4						
	<i>qTGW3.3</i>	3	S03099-S03115	2.8	0.47	10.4						
							<i>qTGW3.1</i>	3	S03145-S03152B	2.94	0.37	9.1
	<i>qTGW4.1</i>	4	S04077A-RM451	3.02	0.39	7.2						
	<i>qTGW4.2</i>	4	RM127-S04129B	3.44	-0.41	7.9						
	<i>qTGW7.1</i>	7	RM248-RM172	2.41	0.30	4.3						
	<i>qTGW10.1</i>	10	RM258-S10058A	3.02	0.36	6.4	<i>qTGW9.1</i>	9	RM108-RM205	3.68	-0.41	11.1
GYP							<i>qGYP1.2</i>	1	RM259-RM600	3.85	1.77	5.6
	<i>qGYP5.1</i>	5	S05014B-RM267	7.74	-2.72	16.0						
							<i>qGYP5.1</i>	5	S05095-RM480	6.95	2.43	10.6
							<i>qGYP6.1</i>	6	S06015-S06018	8.24	-2.86	14.6
							<i>qGYP8.1</i>	8	S08059-S08066	2.29	1.26	2.8
							<i>qGYP10.1</i>	10	S10019-S10026C	3.77	-1.80	5.8
PYP	<i>qPYP1.1</i>	1	S01140-S01143A	3.06	1.50	6.5						
							<i>qPYP6.1</i>	6	RM50-RM527	12.83	4.55	31.7
	<i>qPYP7.1</i>	7	RM234-S07101	3.97	1.69	8.3						
	<i>qPYP8.2</i>	8	S08090-S08106	2.97	1.71	8.5						
							<i>qPYP11.1</i>	11	S11112-S11117B	2.6	1.55	3.7

^a See Table 1 for abbreviations. ^b LOD: logarithm of odds number ^c In the BC1F1s and MPH data set, QTL effects were estimated by the difference between

Suppl Table S4. Comparative studies on the identified heterosis QTLs in two indica-japonica RILs.

Chr.	Source	QTL ^a	Heterosis markers			IRGSP-1.0 ^b		QTARO ^c																			
			Chr.	Flanking markers	LOD ^d	α^e	$R^2(\%)^f$	Start (bp)	Stop (bp)	Pop type	Start (kb)	Stop (kb)	Name	Trait	Parent A	Parent B	Reference										
1	MT	<i>qSN1.1</i>	1	S01022-RM1	15.76	15.96	26.7	4387874	4636870	RIL	1039868	5759851	number of filled grain per panicle	Zhenshan 97B	Miyang 46	Zhang et al. 2000											
										RIL	1405980	5759851	qFLL-1	first leaf length	Zhenshan 97B	Miyang 46	Shen et al. 2003										
										RIL	1405980	5759851	qFLP-1	first leaf perimeter	Zhenshan 97B	Miyang 46	Shen et al. 2003										
										F2	1887524	4635870	gp1a	Grains per panicle	Zhenshan 97B	Minghui 63	Yu et al. 1997										
										BC1F5	3483439	7970819		Root dry weight	Akihikari	IRAT109	Horie et al. 2006										
										BC1F5	3483439	10753685		Root axis length	Akihikari	IRAT109	Horie et al. 2006										
										BC4F4	4424186	4424560	qPN1	Panicle per plant	Guichao 2	Chinese common wild rice (<i>Oryza rufipogon</i> Griff.)	Tian et al. 2006										
										BC2F1	4424186	5941561	ph1.1	Plant height	IR 58025A	O. rufipogon (IC 22015)	Marri et al. 2005										
										RIL	4635793	5759851	qTNSP-1-1	total number of spikelets per panicle	Zhenshan 97B	Miyang 46	Zhang et al. 2002										
										BC1F5	3483439	7970819		Root dry weight	Akihikari	IRAT109	Horie et al. 2006										
										BC1F5	3483439	10753685		Root axis length	Akihikari	IRAT109	Horie et al. 2006										
										1	MT	<i>qGYP1.2</i>	1	RM259-RM600	3.85	1.77	5.6	7446642	9464788	RIL	5094276	7970819	ppp1.1	panicle per plant	Miyang 23	Gihobyee	Cho et al. 2007
RIL	5094276	7970819	spp1.2	spikelets per panicle	Miyang 23	Gihobyee	Cho et al. 2007																				
F2	6273283	9865716	An9	Awmed panicle	Taichung 65	O. meridionalis (Acc. W1625) introgression line	Matsushita et al. 2003																				
F2	5941444	8139063	rf	pollen sterility	IR57596A	IR68077-82-22-2-3R	Sattari et al. 2007																				
RIL	5094276	7970819	prg1.2	percent ripened grain	Miyang 23	Gihobyee	Cho et al. 2007																				
RIL	5759741	3964636		number of filled grains per panicle	Zhenshan 97B	Miyang 46	Zhang et al. 2001																				
RIL	5759741	3964636		number of filled grains per panicle	Zhenshan 97B	Miyang 46	Zhang et al. 2001																				
DT	<i>qTGW1.1</i>	1	RM297-S01140	6.12	0.53	13.4	32100610	34820971	RIL											31822907	33817439	qCC-1	Chlorophyll content	ZYQ8	Jingxi 17	Teng et al. 2004	
DT	<i>qPWP1.1</i>	1	S01140-S01143A	3.06	1.50	6.5	34820971	36282595	F2											28571547	36953571	An10	Awmed panicle	Taichung 65	O. meridionalis (Acc. W1625) introgression line	Matsushita et al. 2003	
1	MT	<i>qCL1.2</i>	1	S01143A-S01147	4.97	2.37	10.5	36282595	38160493											BIL	35712600	39470731	qCh1a	chlorophyll contents	Nipponbare	Kasalath	Yang et al. 2003
																				RIL	36226902	39470731	qLVBTS1-1	Number of large vascular bundles/Total spikelets per panicle	Zhenshan 97A	Minghui 63	Cui et al. 2002
																				RIL	36226902	39470731	qNSB1-1	number of secondary branches per panicle	Zhenshan 97A	Minghui 63	Cui et al. 2002
										RIL	36226902	39470731	qSSBp1-1	number of spikelets on secondary branches per panicle	Zhenshan 97A	Minghui 63	Cui et al. 2002										
										RIL	36226902	39470731	yd1	yield	Zhenshan 97B	Minghui 63	Hua et al. 2002										
										RIL	36226902	41822222	yd1b	yield	Zhenshan 97B	Minghui 63	Xing et al. 2002										
										F2	36657777	40959893		white-berry kernels		CSL(Koshihikari x Kasalath)	Koshihikari	Takeshi et al. 2008									
										RIL	36697294	39534042		spikelet fertility	Zhenshan 97B	Miyang 46	Zhang et al. 2001										
										DHL	36697294	42364623		Penetrated root thickness	IR64	Azucena	Zheng et al. 2000										
										DHL	36697294	42364623	qPL-1	Panicle length	IR64	Azucena	Hittalmani et al. 2002										
										BC2F2	36697294	42364623	pl1.1	Panicle length	Jefferson	O. rufipogon (IRGC 105491)	Thomson et al. 2003										
										1	MT	<i>qGW2.3</i>	3	S03099-S03115	2.8	0.47	10.4	25041462	26658164	DHL	36697294	42364623	sd1	Plant height	IR64	Azucena	Li et al. 2003
BC2F2	15401242	25890781	gw3.1	Grain weight	Jefferson	O. rufipogon (IRGC 105491)	Thomson et al. 2003																				
BC3F2	16196565	25360678	An5(t)	Awless	Asominori	IR24	Kubo et al. 2001																				
CSSL	16196565	25360678	qGL-3	grain length	Asominori	IR24	Wan et al. 2005																				
CSSL	16196565	25360678	qLWR-3	grain length-width ratio	Asominori	IR24	Wan et al. 2005																				
F2	16840989	29580503		Mesocotyl length	Labelle	Black Gora	Redona et al. 1998																				
RIL	18350854	28859103	qSV-3-1	Shoot dry weight	Lemont	Teqing	Zhang et al. 2005																				
F2	23850041	25360678	gw3a	Grain weight, g/1,000	Zhenshan 97A	Minghui 63	Yu et al. 1997																				
DHL	23850041	28999468	Yd3-13	Yield	IR64	Azucena	Yang et al. 2007																				
F2	23850041	29580503		Panicle size	Labelle	Black Gora	Redona et al. 1998																				
F2	23850041	31298638		Coleoptile length	Labelle	Black Gora	Redona et al. 1998																				
1	MT	<i>qGW2.1</i>	3	S03145-S03152B	2.94	0.37	9.1	32235745	33938972											RIL	BC2F1	25891087	27899169	QFlw3	flag-leaf width	Lemont	Teqing
										DHL	31082622	35089139	qPES-3	Percentage of exerted stigmas	Zaiyeqing 8	Jingxi 17	Li et al. 2003										
										RIL	31082622	37254205		grains per plant	Lemont	Teqing	Luo et al. 2007										
										RIL	31082622	37254205	QFl3	flag leaf length	Lemont	Teqing	Mei et al. 2003										
										RIL	31082622	37254205	QSn3	spikelet number per panicle	Lemont	Teqing	Mei et al. 2003										
										DT	<i>qGW4.1</i>	4	S04077A-RM451	3.02	0.39	7.2	25177554	28571472	RIL	21645173	34244441	gpp4	grains per panicle	9024	LH422	Xiao et al. 1996	
										DT	<i>qPL4.1</i>	4	S04077A-RM451	5.38	0.45	9.4	25177554	28571472	RIL	22859833	27441315	qCh1a-1	Chlorophyll a	Zhenshan 97A	Minghui 63	Cui et al. 2002	
										RIL	22859833	27441315	qCh1a-1	Total Chlorophyll	Zhenshan 97A	Minghui 63	Cui et al. 2002										
										RIL	22859833	27441315	qPH1-4-1	plant height at 3SDAT	Zhenshan 97	Minghui 63	Cui et al. 2004										
										CSSL	25273817	25274481	qGP4	grain number	Asominori	IR24	Fan et al. 2005										
										BC2F2	25316661	33033092	ps4.1	Percent seed set	Jefferson	O. rufipogon (IRGC 105491)	Thomson et al. 2003										
										RIL	27441052	27441315	qSLTS4-1	Area of -2nd leaf/Total spikelets per panicle	Zhenshan 97	Minghui 63	Cui et al. 2003										
RIL	27941367	30762314		Maximum new root length Average	Otomemochi	Yumehotatamochi	Ikedai et al. 2006																				
1	MT	<i>qPL4.2</i>	4	S04120-S04128	7.58	0.57	11.7	33821212	34789152	F2	32246485	34252531		average primary branch number per panicle per plant	Palawan	IR42	Wu et al. 1996										
										F2	32246485	34252531	QGN4	grain number per panicle	Lemont	Teqing	Li et al. 1997										
										RIL	34553977	34561100	qSPR4	spreading of panicle at maturity stage	A58	W107 (o. rufipogon)	Onishi et al. 2007										
										F2	34553977	35281498	Sh3	seed shattering	Taichung 65	n Griff. (Acc. IRGC105715) intro	Nagai et al. 2002										
										F2	32246485	34252531	np4	Number of panicles per plant	Waiyin 2	CB	Lin et al. 1995										
										F2, F3	32246485	35283358		number of productive panicles per plant	Tesanaï	CB	Zhuang et al. 1996										
										F2	34553977	35281498	Sh3-gla	seed shattering	Taichung 65	glaberrima (ACC IRGC10403)	Nagai et al. 2002										
										RIL	21645173	34244441	gpp4	grains per panicle	9024	LH422	Xiao et al. 1996										
										F2	32246485	34252531	tgw4	1000-grain weight	Tesanaï 2	CB	Lin et al. 1995										
										F2	32246485	34252531	np4	Number of panicles/plant	IR42	CB	Lin et al. 1996										
										F2	31436862	34252531	QGWp4	chlorophyll content	Palawan	Teqing	Wu et al. 1996										
										F2	32246485	34252531		grain weight per panicle	Lemont	Teqing	Li et al. 1997										
1	DT	<i>qTGW4.2</i>	4	RM127-S04129B	3.44	-0.41	7.9	34714843	35012117	F2	34553977	35281498	Sh3	seed shattering	Taichung 65	n Griff. (Acc. IRGC105715) intro	Nagai et al. 2002										
										F2	34553977	35281498	Sh3-gla	seed shattering	Taichung 65	glaberrima (ACC IRGC10403)	Nagai et al. 2002										
1	DT	<i>qGYP5.1</i>	5	S05014B-RM267	7.74	-2.72	16.0	1515603	2881479	BIL	238143	3285751	CORN5	the ratio of Rubisco to total leaf N content at 5days after heading	Nipponbare	Kasalath	Ishimaru et al. 2001										
										BC1F1	836248	3445769	qSF5.2	hybrid fertility	Ipumbuyo	Dasanbyeo	Refinur et al. 2012										
										DHL	2069338	2760413		seminal root length(upland)	Azucena	Azucena	Zheng et al. 2006										
										DHL	2069338	2760413	PH5-1	plant height	IR64	Azucena	Yan et al. 1998										
1	MT	<i>qPN5.2</i>	5	RM163-S05087	3.27	0.38	7.5	19251933	21237962	RIL	18760746	19982059	dcs5	degree of chlorophyll content of the second leaf at the heading date	SNU-SG1	Miyang 23	Yoo et al. 2007										
										F2, F3	18944261	21678878		1000 grain weight	Tesanaï	CB	Zhuang et al. 1996										
										F2	18944261	29429411	QPIS	Spikelet density	Lemont	Teqing	Li et al. 1998										
										DHL	19258755	28755091		grain width	Reho	Yamadanshiki	Yoshida et al. 2002										
										DHL	19258755	28755091		grain thickness	Reho	Yamadanshiki	Yoshida et al. 2002										
										F2	20217134	20220313	SH5-glum	seed shattering	Taichung 65	meridionalis introgression line	Sanchez et al. 2002										
										F2	18944261	29429411	QPIS	Spikelet density	Lemont	Teqing	Li et al. 2008										
										DHL	19258755	28755091		grain width	Reho	Yamadanshiki	Yoshida et al. 2002										
										DHL	19258755	28755091		grain thickness	Reho	Yamadanshiki	Yoshida et al. 2002										
										F2	23591602	29429411	QFla5	flag leaf angle	Lemont	Teqing	Li et al. 1999										
										F2	23591602	29429411	Eja	elongated uppermost internode	307T	Zhenshan 97	Xu et al. 2004										
										RIL	24086072	27457701	qRGR-5	Relative germination rate(RGR)	Zhenshan 97	IRAT109	You et al. 2006										
RIL	26972040	27050414	qm5-4	mesocotyl length	Zhenshan 97B	Miyang 46	Ouyang et al. 2005																				
RIL	27055983	27486349	qRDW5-1	Root dry weight	Zhenshan 97	Minghui 63	Cui et al. 2002																				
RIL	27055983	27486349	qRAS-1	Root activity	Zhenshan 97	Minghui 63	Cui et al. 2002																				
RIL	27055983	27486349	qGRS-1	Germination rate	Zhenshan 97	Minghui 63	Cui et al. 2002																				
1	MT	<i>qGYP6.1</i>	6	S06015-S06018	8.24	-2.86	14.6	4194946	4738670	BC4F2	2684129	4352379	Lhd1(t)	Flowering	RGC Acc. No. 105661	Taichung 65	Sanchez et al. 2002										
										BC1F1	2853061	4303488	d4	Dwarf	H125	Kasalath	Maekawa et al. 2005										

										RIL	3168374	4234201	qSL6-6	second leaf length	Zhenshan 978	Miyang 46	Shen et al. 2003
										RIL	3459492	6023472		total number of spikelets per panicle	Zhenshan 978	Miyang 46	Zhuang et al. 2001
										RIL	3459492	6023472		Grain yield	Zhenshan 978	Miyang 46	Zhuang et al. 2001
										RIL	3459492	6023472		spikelet fertility	Zhenshan 978	Miyang 46	Zhuang et al. 2001
											4200251	4201852	DPL2	Sterility			Mizuta et al. 2010
										RIL	4212289	4899565	qNP-6	number of panicles per plant	Zhenshan 978	Miyang 46	Zhuang et al. 2002
										BC1F1	4234080	5096667	qLW-6	Grain shape	Balilla	NTH	Yan et al. 2003
										BC1F2	4234080	5096667	qSP1F6	spikelet fertility	Balilla	NTH	Yan et al. 2003
										BC1F2	4234080	6399814	qRCR1-6	Reduction in chlorophyll content	IR36	Nekken2	Abdelkhaliq et al. 2005
										BC1F2	4234080	6399814	qRCR1-6	Reduction in chlorophyll content	IR36	Nekken2	Abdelkhaliq et al. 2005
										F2	6023974	9537572	tns6	Total number of spikelets/panicle	Tesana 2	CB	Lin et al. 1995
										F2	6023974	9537572	sd6	Spikelet density	Tesana 2	CB	Lin et al. 1995
										F2	6023974	9537572	tns6	Total number of spikelets/panicle	Waiyin 2	CB	Lin et al. 1996
										DHL	6230045	7592308		the spikelet number of a panicle	Zhenshan 97	Wuyujing 2	Jiang et al. 2004
											6283432	7177183	sp6f	Spikelet fertility	02428/Nanjing12	Balilla	Song et al. 2005
											6283432	9284248	S5	spikelet fertility	Nanjing11	Duiai	Yan et al. 2000
										RIL	6521345	6927968	qNPB6-1	number of primary branches per panicle	Zhenshan 97	Minghui 63	Cui et al. 2002
										RIL	6720901	8066362	qPSP-6	photoperiod sensitive phase	O. rufipogon/W1944	Pei-Khu	Cai and Morishima 1998
										RIL	6927624	20691040	gp6	Grains/panicle	Zhenshan 97	Minghui 63	Hua et al. 2002
										RIL	6927624	20691040	qLV8L6-1	number of large vascular bundles/Area of -2n	Zhenshan 97	Minghui 63	Cui et al. 2003
										RIL	6927624	20691040	qR56-1	Reducing sugar content of seedling	Zhenshan 97	Minghui 63	Cui et al. 2002
										RIL	6927624	20691040	qGY6-1	Grain yield (t/ha)	Zhenshan 97	Minghui 63	Cui et al. 2003
										RIL	6927624	20691040	qS16-1	Area of -2nd leaf (cm ²)	Zhenshan 97	Minghui 63	Cui et al. 2003
										RIL	6927624	20691040	qTN2-6-1	tiller number at maturity	Zhenshan 97	Minghui 63	Cui et al. 2004
										RIL	6927624	20691040	qPH2-6-1	plant height at maturity	Zhenshan 97	Minghui 63	Cui et al. 2004
										RIL	6927624	20691040	qHD-6-1	heading date	Zhenshan 97	Minghui 63	Cui et al. 2004
										RIL	6927624	29906021	ph6	plant height	9024	LH422	Xiao et al. 1996
										DHL	7592048	8751554		the number of grains per panicle (GP)	Zhenshan 97	Wuyujing 2	Jiang et al. 2004
										DHL	7592048	8751554		the effective tiller percentage per plant(TP)	Zhenshan 97	Wuyujing 2	Jiang et al. 2004
										BIL	8054255	8066362	qN6-6	6th internode length	Koshihikari	Kasalath	Yamamoto et al. 2001
										BIL	8054255	8066362	qDTH-6	days-to-heading	Koshihikari	Kasalath	Yamamoto et al. 2001
											8054255	8066362	Su-Se-1(t)	independent photoperiod-sensitive suppression	Asominori	IR24	Yu et al. 2005
										RIL	8751256	12402719	qNFGP-6	number of filled grains per panicle	Zhong 156	Gumei 2	Zhuang et al. 2001
										RIL	9282143	17933378	qSPN-6	Number of spikelets per panicle	ZYQ8	Jingxi 17	He et al. 2001
										DHL	9282143	17933378	gw-6	1000-grain weight	Zaiyeqing 8	Jingxi 17	Liu et al. 1996
										F2	2349842	17535483	gp7a	Grains/panicle	Zhenshan 97	Minghui 63	Li et al. 2000
										F2	2349842	17535483	gw7	1000 Grain weight	Zhenshan 97	Minghui 63	Li et al. 2000
										F2	2349842	17535483	yd7a	Yield per plant	Zhenshan 97	Minghui 63	Li et al. 2000
										F2	2349842	17535483	tp7b	Tillers/plant	Zhenshan 97	Minghui 63	Li et al. 2000
										F2	4606397	6812968	qHD7	heading date	IR64	Azucena	Li et al. 2003
										DHL	4606397	16264722	qh7	length of the heading period	Nipponbare	Kasalath	Ishimaru and Kashiwagi, 2004
										BIL	4606397	17535483		heading date	Hoshiyounme	Kasalath	Nonoue et al. 2008
										F2	5512628	5512754	dth7.1	Days to heading	IR64	O. rufipogon (IRGC 105491)	Septiningsih et al. 2003
										BC2F2	5512628	19619933	qh7d-7	heading date	USSR5	N22	Jiang et al. 2007
										F2	5884863	6812968	pms1	photoperiod-sensitive genetic male sterility	Minghui 63	320015	Liu et al. 2001
											8358000	11394315	Hd4	days to heading	Nipponbare	NIL (Hd4)	Lin et al. 2003
										BC3F2	8910286	11394315	qSSP7	number of spikelets per panicle	Zhenshan 97	Minghui 63	Xing et al. 2008
										RIL	11391449	11394315	qhD-1	days to heading	A58	W107 (o. rufipogon)	Onishi et al. 2007
										RIL	18481047	25763098	QR17	leaf rolling	Lemont	Teqing	Xu et al. 1999
										RIL	19354374	26133946	qSDS-7-2	Seed dormancy	EM93-1	SS18-2	Gu et al. 2004
										BC1F1	24746657	26133946	7-3	seed maximum root length,number of roots	Bala	Azucena	Price et al. 2002
										BC1F1	26133781	30222602	qPLL7	Pollen fertility	Balilla	NTH	Yan et al. 2003
										F3	26313662	27191049	hd7a	Heading date	Zhenshan 97	Minghui 63	Yu et al. 2002
										BC2F2	27188091	30322668	S23(t)	Sterility	RGCC Acc. No. 10566	Taichung 65	Sobrizal et al. 2000
										BIL	27391198	29608218	qDTH-7	day to heading	Hayamasari	Italica Livorno	Fujino and Sekiguchi, 2005
										F2	360155	4446617	qP8L	Panicle length	98SQ1496	Koshihikari	Miyata et al. 2007
										F2	4337117	4335434	RH8	heading date and heterosis		Li et al. 2016	
										F2	360155	4446617	qDTH8	Days to heading	98SQ1496	Koshihikari	Miyata et al. 2007
										F2	4105519	5425677		Grain weight	Black Gora	LH422	Redona et al. 1998
										RIL	4105519	25684949	dth8	Days to heading	9024	Xiao et al. 1996	
										BC	4444681	4446617	Hd5	Days to heading	Nipponbare	Hd5-NIL	Lin et al. 2003
										F2	5421262	7568613	ntf8	Number of first branches per panicle	Waiyin 2	CB	Lin et al. 1995
										F2	5421262	17528755	hd8	heading date	Tesani 2	CB	Lin et al. 1995
										RIL	4105519	25684949	dth8	Days to heading	9024	LH422	Xiao et al. 1996
										F2	5421262	17528755	hd8	heading date	Tesani 2	CB	Lin et al. 1995
										BIL	19395396	24277473		Rubisco protein content at 5days after heading	Nipponbare	Kasalath	Ishimaru et al. 2001
										BC3F2	20740779	24569536	qPN-8	Panicles per plant	Miyang 23	Oryza glaberrima (accession IRGC No. 103544)	Suh et al. 2005
										RIL	24274675	27825769	8-4	root to shoot ratio,maximum root length	Bala	Azucena	Price et al. 2002
											25362546	25366701	WFP	panicle branching, Tillering, filled grain weight per plant	ST12	CB	Miura et al. 2010
										F2, F3	25681306	27825769		root to shoot ratio,maximum root length	Bala	Azucena	Zhuang et al. 1996
										RIL	24274675	27825769	8-4	root to shoot ratio,maximum root length	Bala	Azucena	Price et al. 2002
										F2, F3	25681306	27825769		filled grain weight per plant	Tesani	CB	Zhuang et al. 1996
										RIL	26582935	28166213	qRGV-8	Relative germination vigor	Zhenshan 97	IRAT109	You et al. 2006
										DH	1159676	11367960		Maximum root length	ZYQ8	Jingxi 17	Xu et al. 2001
										BC1F2	18374393	19465043	qGL9	Grain length	98SQ1496	Koshihikari	Miyata et al. 2007
										DHL	18374393	20828859	qCCAI-9	f chlorophyll content 25 days after flowering	IR36	Nekken2	Abdelkhaliq et al. 2005
										F2	19168716	20079157	ta9	tiller angle	IR64	Azucena	Yan et al. 1999
										F2	19168716	21314308	Ta1	tiller, leaf and flag leaf angles	Lemont	Teqing	Li et al. 1998
										F2	19168716	21314308	Ta	tillering angle	Lemont	Teqing	Li et al. 1999
										RIL	19464779	22021406	gy9	grain yield	Dasanbyeol	TR22183	Cho et al. 2007
										BC2F1	19464779	2352932	ylpd9.1	Yield per plant	IR 58025A	O. rufipogon (IC 22015)	Marri et al. 2005
										F2	20442345	20828859	qMXN-9	number of late metaxylem vessels			

Fujino, K., and Sekiguchi, H. (2005). Identification of QTLs Conferring Genetic Variation for Heading Date among Rice Varieties at the Northern-limit of Rice Cultivation. *Breeding Science* 55, 141-146.

Fukuta, Y., and Yagi, T. (1998). Mapping of a Shattering Resistance Gene in a Mutant Line SR-5 Seeded from an India Rice Variety, Nan-jing 11. *Breeding Science* 48, 345-348.

Gu, X.-Y., Kianian, S.F., and Foley, M.E. (2004). Multiple Loci and Epistases Control Genotype Variation for Seed Dormancy in Weedy Rice (*Oryza sativa*). *Genetics* 166, 1503-1516.

He, P., Li, J.Z., Zheng, X.W., Shen, L.S., Lu, C.F., Chen, Y., and Zhu, L.H. (2001). Comparison of Molecular Linkage Maps and Agronomic Trait Loci between DH and RIL Populations Derived from the Same Rice Cross. *Crop Sci.* 41, 1240-1246.

Hittalmani, S., Huang, N., Courtis, B., Venuprasad, R., Shashidhar, H.E., Zhuang, J.Y., Zheng, K.L., Liu, G.F., Wang, G.C., Sidhu, J.S., Srinivasanayak, S., Singh, V.P., Bagal, P.G., Prasanna, H.C., McLaren, G., and Khush, G.S. (2003). Identification of QTL for growth- and grain yield-related traits in rice across nine locations of Asia. *Theor. Appl. Genet.* 107, 679-690.

Hittalmani, S., Shashidhar, H.E., Bagal, P.G., Huang, N., Sidhu, J.S., Singh, V.P., and Khush, G.S. (2002). Molecular mapping of quantitative trait loci for plant growth, yield and yield related traits across three diverse locations in a doubled haploid rice population. *Euphytica* 125, 207-214.

Hori, H., Nemoto, K., Miyamoto, N., and Harada, J. (2006). Quantitative trait loci for adventitious and lateral roots in rice. *Plant Breeding* 125, 198-200.

Hua, J.P., Ying, Y.Z., Xu, C.G., Sun, X.J., Yu, S.B., and Zhang, Q. (2002). Genetic Dissection of an Elite Rice Hybrid Revealed That Heterozygotes are Not Always Advantageous for Performance. *Genetics* 162, 1885-1895.

Huang, N., Parco, A., Mew, T., Magarathay, G., McCouch, S., Guiderdoni, E., Xu, J., Sathuthi, P., Angeles, E.R., and Khush, G.S. (1997). RFLP mapping of isozymes, RAPD and QTLs for grain shape, brown planthopper resistance in a doubled haploid rice population. *Molecular Breeding* 3, 105-113.

Iwata, H., Kamoshita, A., and Marabe, T. (2006). Genetic analysis of rooting ability of transplanted rice (*Oryza sativa* L.) under different water conditions. *J. Exp. Bot.* e1162.

Ishimaru, K., and Kashiwagi, T. (2004). Identification of a locus for asynchronous heading in rice, *Oryza sativa* L. *Euphytica* 139, 141-145.

Ishimaru, K., Kobayashi, N., Ono, K., Yano, M., and Ohtsugi, R. (2001). Are contents of Rubisco, soluble protein and nitrogen in flag leaves of rice controlled by the same genetics? *J. Exp. Bot.* 52, 1827-1833.

Jiang, G.H., Xu, C.G., Li, X.H., and He, Y.Q. (2004). Characterization of the genetic basis for yield and its component traits of rice revealed by doubled haploid population. *Yi Chuan Xue Bao* 31, 63-72.

Jiang, L., Xu, J., Wei, X., Wang, S., Tang, J., Zhai, H., and Wan, J. (2007). The inheritance of early heading in the rice variety USSR5. *J. Genet. Genomes* 34, 46-55.

Zhang, L., Zhang, W., Xia, Z., Jiang, G., Qian, Q., Li, A., Cheng, Z., Zhu, L., Mao, L., and Zhai, W. (2007). A paracentric inversion suppresses genetic recombination at the F0N3 locus with breakpoints corresponding to sequence gaps on rice chromosome 11L. *Mol. Genet. Genomics* 277, 263-272.

Kamohita, A., Zhang, J., Siopongco, J., Sankarung, S., Nguyen, H.T., and Wade, L.J. (2002). Effects of Phenotyping Environment on Identification of Quantitative Trait Loci for Rice Root Morphology under Anaerobic Conditions. *Crop Sci.* 42, 255-265.

Kubo, T., Takano-Kai, N., and Yoshimura, A. (2001). RFLP mapping of genes for long kernel and awn on chromosome 3 in rice. *Rice Genetics Newsletter* 18, 26-28.

Li, D.T., Zhou, S.S., Jiang, L., and Wan, J.M. (2005). Mapping for a new locus causing hybrid sterility in a China landrace (*Oryza sativa* L.). *Rice Genetics Newsletter* 22, 20-.

Li, W.H., Dong, G.J., Hu, X.M., Teng, S., Guo, L.B., Zheng, D.L., and Qian, Q. (2003). QTL analysis for percentage of exerted stigma in rice (*Oryza sativa* L.). *Yi Chuan Xue Bao* 30, 637-640.

Li, Z., Paterson, A.H., Pinson, S.R.M., and Khush, G.S. (1998). A major gene, Tal and QTLs affecting tiller and leaf angles in rice. *Rice Genetics Newsletter* 15, 154-156.

Li, Z., Paterson, A., Pinson, S., and Stansel, J. (1999). RFLP facilitated analysis of tiller and leaf angles in rice (*Oryza sativa* L.). *Euphytica* 109, 79-84.

Li, Z., Pinson, S.R., Park, W.D., Paterson, A.H., and Stansel, J.W. (1997). Epistasis for three grain yield components in rice (*Oryza sativa* L.). *Genetics* 145, 453-465.

Li, Z., Pinson, S.R., Stansel, J.W., and Paterson, A.H. (1998). Genetic dissection of the source-sink relationship affecting fertility and yield in rice (shape *Oryza sativa* L.). *Molecular Breeding* 4, 419-426.

Li, Z.K., Yu, S.B., Lafitte, H.R., Huang, N., Courtis, B., Hittalmani, S., Vijayakumar, C.H., Liu, G.F., Wang, G.C., Shashidhar, H.E., Zhuang, J.Y., Zheng, K.L., Singh, V.P., Sidhu, J.S., Srinivasanayak, S., and Khush, G.S. (2003). QTL x environment interactions in rice. I. heading date and plant height. *Theor. Appl. Genet.* 106, 141-153.

Li, J.X., Yu, S.B., Xu, C.G., Tan, Y.F., Gao, Y.J., Li, X.H., and Zhang, Q. (2000). Analyzing quantitative trait loci for yield using a vegetatively replicated F2 population from a cross between the parents of an elite rice hybrid. *TAG Theoretical and Applied Genetics* 101, 248-254.

Lin, H., Liang, Z.-W., Sasaki, T., and Yano, M. (2003). Fine Mapping and Characterization of Quantitative Trait Loci Hd4 and Hd5 Controlling Heading Date in Rice. *Breeding Science* 53, 51-59.

Lin, H.X., Qian, H.R., Zhuang, J.Y., Lu, J., Min, S.K., M.Xiong, Z., Huang, N., and Zheng, K.L. (1995). Interval mapping of QTLs for yield and other related characters in rice. *Rice Genetics Newsletter* 12, 251-253.

Lin, H.X., Qian, H.R., Zhuang, J.Y., Xiong, J.L.Z.M., Min, S.K., Huang, N., and Zheng, K.L. (1995). RFLP mapping of major genes and minor genes for heading date in rice (*Oryza sativa* L.). *TAG Theoretical and Applied Genetics* 92, 920-927.

Lin, H.X., Qian, H.R., Zhuang, J.Y., Lu, J., Min, S.K., Xiong, Z.M., Huang, N., and Zheng, K.L. (1996). RFLP mapping of QTLs for agronomic traits of rice across environments using a doubled haploid population. *TAG Theoretical and Applied Genetics* 93, 1211-1217.

Liu, N., Shan, Y., Wang, F.P., Xu, C.G., Peng, K.M., Li, X.H., and Zhang, Q. (2001). Identification of an 85-kb DNA fragment containing *pnsl1*, a locus for photoperiod-sensitive gene male sterility in rice. *Mol. Genet. Genomics* 266, 271-275.

Lu, C., Shen, L., Tan, Z., Xu, Y., He, P., and Zhu, L. (1996). Comparative mapping of QTLs for agronomic traits of rice across environments using a doubled haploid population. *TAG Theoretical and Applied Genetics* 93, 1211-1217.

Luo, L.J., Li, Z.K., Mei, H.W., Shi, Q.Y., Tabien, R., Zhong, D.B., Ying, C.S., Stansel, J.W., Khush, G.S., and Paterson, A.H. (2001). Overdominant Epistatic Loci Are the Primary Genetic Basis of Inbreeding Depression and Heterosis in Rice. II. Grain Yield Components. *Genetics* 158, 1755-1771.

Maekawa, M., Takamura, I., Ahmed, N., and Kyozuka, J. (2005). Bunketsu-uo-1, one of the Thirring Dwarfs, is Controlled by a Single Recessive Gene in Rice (*Oryza sativa* L.). *Breeding Science* 55, 193-196.

Mari, P.R., Sarla, N., Reddy, L.V., and Siddiq, E.A. (2005). Identification and mapping of yield related QTLs from an Indian accession of rice *Oryza rufipogon*. *BMC Genet.* 6, 33.

Matsushita, S., Kurakazu, T., Sobriazi, Doi, K., and Yoshimura, A. (2003). Mapping of genes for awn in rice using *Oryza meridionalis* introgression lines. *Rice Genetics Newsletter* 20, 17-18.

Mei, H.W., Luo, L.J., Ying, C.S., Wang, Y.P., Yu, X.Q., Guo, L.B., Paterson, A.H., and Li, Z.K. (2003). Gene actions of QTLs affecting several agronomic traits resolved in a recombinant inbred rice population and two testcross populations. *Theor. Appl. Genet.* 107, 89-101.

Mei, H.W., Li, Z.K., Shi, Q.Y., Guo, L.B., Wang, Y.P., Yu, X.Q., Ying, C.S., and Luo, L.J. (2005). Gene actions of QTLs affecting several agronomic traits resolved in a recombinant inbred rice population and two backcross populations. *Theor. Appl. Genet.* 110, 649-659.

Miura, K., Ikeda, M., Matsubara, A., Song, X.-J., et al. (2010) OsSPL14 promotes panicle branching and higher grain productivity in rice. *Nature Genet.* 42:545-549.

Miyata, M., Yamamoto, T., Komori, T., and Nitta, N. (2007). Marker-assisted selection and evaluation of the QTL for stigma exertion under japonica rice genetic background. *Theor. Appl. Genet.* 114, 539-548.

Miyamoto, N., Goto, Y., Matsui, M., Ueki, Y., Morita, M., and Nemoto, K. (2004). Quantitative trait loci for phytochrome and tillering in rice. *Theor. Appl. Genet.* 109, 700-706.

Mizuta, Y., Hanushima, Y., Kurata, N. (2010) Rice pollen hybrid incompatibility caused by reciprocal gene loss of duplicated genes. *PNAS* 107(47): 20417-20422.

Nagai, Y.S., Sobriazi, Sanchez, P.L., Kurakazu, T., Doi, K., and Yoshimura, A. (2002). Sh3, a gene for seed shattering, commonly found in wild rices. *Rice Genetics Newsletter* 19, 74-76.

Nonoue, Y., Fujino, K., Hirayama, Y., Yamanouchi, U., Lin, S.Y., and Yano, M. (2008). Detection of quantitative trait loci controlling extremely early heading in rice. *Theor. Appl. Genet.* 116, 715-722.

Ochiai, K., Uemura, S., Shimizu, A., Okumoto, Y., and Matoh, T. (2008). Boron toxicity in rice (*Oryza sativa* L.). I. Quantitative trait locus (QTL) analysis of tolerance to boron toxicity. *Theor. Appl. Genet.* 117, 125-133.

Onishi, K., Horiuchi, Y., Shigeh-Oka, N., Takagi, K., Ichikawa, N., Maruoka, M., and Sano, Y. (2007). A QTL Cluster for Plant Architecture and Its Ecological Significance in Asian Wild Rice. *Breeding Science* 57, 7-16.

Ouyang, Y.H., Zhang, Q.Y., Zhang, K.Q., Yu, S.M., Zhuang, J.Y., Jin, Q.Y., and Cheng, S.H. (2005). QTL mapping and interaction analysis of genotype x environment (Fe2+ concentrations) for mesocotyl length in rice (*Oryza sativa* L.). *Yi Chuan Xue Bao* 32, 712-718.

Price, A.H., Steele, K.A., Moore, B.J., and Jones, R.G. (2002). Upland rice grown in soil-filled channels and exposed to contrasting water-deficit regimes: II. Mapping quantitative trait loci for root morphology and distribution. *Field Crops Research* 76, 25-43.

Qu, Y., Mu, P., Zhang, H., Chen, C.Y., Gao, Y., Tian, Y., Wen, F., and Li, Z. (2008). Mapping QTLs of root morphological traits at different growth stages in rice. *Genetics* 133, 187-200.

Reddy, E.D., and Mackill, D.J. (1998). Quantitative trait locus analysis for rice panicle and grain characteristics. *TAG Theoretical and Applied Genetics* 96, 957-963.

Refinur, Chin, J.H., et al. (2012) QTLs for hybrid fertility and their association with female and male sterility in rice. *Genes and Genomics* 34: 355-365.

Sanchez, P.L., Sobriazi, Ikeda, K., Yashi, H., and Yoshimura, A. (2000). Linkage analysis of late heading genes using *Oryza glumepatula* introgression lines. *Rice Genetics Newsletter* 17, 46-48.

Sanchez, P.L., Kurakazu, T., Hirata, C., Sobriazi, and Yoshimura, A. (2002). Identification and mapping of seed shattering genes using introgression lines from wild rice species. *Rice Genetics Newsletter* 19, 78-80.

Sattari, M., Khatresani, A., Geronzi, G., and Virmani, S. (2007). Comparative genetic analysis and molecular mapping of fertility restoration genes for WA, Dissi, and Gambiza cytoplasmic male sterility systems in rice. *Euphytica*.

Septimings, E.M., Prasejyano, J., Lubis, E., Tai, T.H., Tjabyant, S., Moeljowiro, S., and McCouch, S.R. (2003). Identification of quantitative trait loci for yield and yield components in an advanced backcross population derived from the *Oryza sativa* ssp. *ir64* and the wild relative *O. rufipogon*. *Theor. Appl. Genet.* 107, 1419-1432.

Shen, B., Zhuang, J.Y., Zhang, K.Q., Xia, Q.Q., Sheng, C.X., and Zheng, K.L. (2003). QTLs mapping of leaf traits and root vitality in a recombinant inbred line population of rice. *Yi Chuan Xue Bao* 30, 1133-1139.

Sobriazi, Matsubara, Y., Sanchez, P.L., Ikeda, K., and Yoshimura, A. (2000). Mapping of F3 pollen semi-sterility gene found in backcross progeny of *Oryza sativa* L. and *Oryza glumepatula* Steud. *Rice Genetics Newsletter* 17, 61-63.

Song, X., Qiu, S.Q., Xu, C.G., Li, X.H., and Zhang, Q. (2005). Genetic dissection of embryo sac fertility, pollen fertility, and their contributions to spikelet fertility in interspecific hybrids in rice. *Theor. Appl. Genet.* 110, 205-211.

Suh, J.P., Ahn, S.N., Cho, Y.C., Kang, K.H., Choi, I.S., Kim, Y.G., Suh, G.H., and Hong, H.C. (2005). Mapping of QTLs for yield traits using an advanced backcross population from a cross between *Oryza sativa* and *O. glaberrima*. *Korean J. Breed.* 37, 214-220.

陈松志, 山本寿孝, 吴树群, 和藤原康彦. (2008). 染色体定位图数据库构建. *植物分子生物学研究* 10, 91-99.

Tan, X.L., Vanavichit, A., Amornsilpa, S., and Tragoonsum, S. (1998). Genetic analysis of rice CMS-WA fertility restoration based on QTL mapping. *TAG Theoretical and Applied Genetics* 97, 994-999.

Tang, S., Qian, Q., Zeng, D., Kunihiro, Y., Fujimoto, K., Huang, D., and Zhu, L. (2004). QTL analysis of leaf photosynthetic rate and related physiological traits in rice (*Oryza sativa* L.). *Euphytica* 135, 1-7.

Thomson, M.J., Tai, T.H., McClung, A.M., Lai, X.H., Hinga, M.B., Lobos, K.B., Xu, Y., Martinez, C.P., and McCouch, S.R. (2003). Mapping quantitative trait loci for yield, yield components and morphological traits in an advanced backcross population between *Oryza rufipogon* and the *Oryza sativa* cultivar Jefferson. *Theor. Appl. Genet.* 107, 479-493.

Tian, F., Li, D.J., Fu, Q., Zhu, Z.F., Fu, Y.C., Wang, X.K., and Sun, C.Q. (2006). Construction of introgression lines carrying wild rice (*Oryza rufipogon* Griff.) segments in cultivated rice (*Oryza sativa* L.) background and characterization of introgressed segments associated with yield-related traits. *Theor. Appl. Genet.* 112, 570-580.

Uga, Y., Okuno, K., and Yano, M. (2000). QTLs underlying natural variation in staminal and xylem structures of rice root. *Breeding Science* 58, 7-14.

Wada, T., Uchimura, Y., Ogata, T., Tsubone, M., and Matsue, Y. (2006). Mapping of QTLs for Physicochemical Properties in Japonica Rice. *Breeding Science* 56, 253-260.

Wan, X.Y., Wan, J.M., Wang, J.F., Jiang, L., B., J.C., Wang, C.M., and Zhai, Q.Q. (2005). Stability of QTLs for rice grain dimension and endosperm chalkiness characteristics across eight environments. *Theor. Appl. Genet.* 110, 1334-1346.

Wu, P., Zhang, G., and Huang, N. (1996). Identification of QTLs controlling quantitative characters in rice using RFLP markers. *Euphytica* 98, 349-354.

Xiao, J., Li, J., Yuan, L., and Tanksley, S.D. (1996). Identification of QTLs affecting traits of agronomic importance in a recombinant inbred population derived from a subspecific rice cross. *TAG Theoretical and Applied Genetics* 92, 230-244.

Xing, Y.Z., Xu, C.G., Hua, J.P., and Tan, Y.F. (2001). Analysis of QTL x environment interaction for rice panicle characteristics. *Yi Chuan Xue Bao* 28, 439-446.

Xing, Z., Tan, H., Hua, P., Sun, L., Xu, G., and Zhang, Q. (2002). Characterization of the main effects, epistatic effects and their environmental interactions of QTLs on the genetic basis of yield traits in rice. *Theor. Appl. Genet.* 105, 248-257.

Yong, Y.Z., Tang, W.J., Xue, W.Y., Xu, C.G., and Zhang, Q. (2002). Fine mapping of a major quantitative trait loci, qSSP7, controlling the number of spikelets per panicle as a single Mendelian factor in rice. *Theor. Appl. Genet.* 116, 789-796.

Xu, J.C., Li, J.Z., Zheng, X.W., Zou, L.X., and Zhu, L.H. (2001). QTL mapping of the root traits in rice seedling. *Yi Chuan Xue Bao* 28, 433-438.

Xu, J.L., Zhong, D.B., Yu, S.B., Luo, L.J., and Li, Z.K. (1999). QTLs affecting leaf rolling and folding in rice. *Rice Genetics Newsletter* 16, 51-53.

Xu, K., and Mackill, D.J. (1995). RAPD and RFLP mapping of a submergence tolerance locus in rice. *Rice Genetics Newsletter* 12, 244-245.

Xu, Y.H., Zhu, Y.Y., Zhou, H.C., Li, Q., Sun, Z.X., Liu, Y.G., Lin, H.X., and He, Z.H. (2004). Identification of a 98-kb DNA segment containing the rice *Eui* gene controlling uppermost internode elongation, and construction of a TAC transgene library. *Mol. Genet. Genomics* 272, 149-155.

Yamagishi, J., Miyamoto, N., Hirotsu, S., Laza, R.C., and Nemoto, K. (2004). QTLs for branching, foret formation, and pre-flowering abort of rice panicle in a temperate japonica x tropical japonica cross. *Theor. Appl. Genet.* 109, 1555-1561.

Yamamoto, T., Sasaki, T., and Yano, M. (1997). Genetic Analysis of Spreading Stub Using indica/Japonica Backcrossed Progenies in Rice. *Breeding Science* 47, 141-144.

Yamamoto, T., Taguchi-Shiobara, F., Ueki, Y., Sasaki, T., and Yano, M. (2001). Mapping Quantitative Trait Loci for Days-to-heading, and Culm, Panicle and Internode Lengths in a BC1F3 Population Using an Elite Rice Variety, Koshihikari, as the Recurrent Parent. *Breeding Science* 51, 63-71.

Yan, C.J., Liang, G.H., Zhu, L.H., and Gu, M.H. (2000). RFLP analysis on wide compatibility genes in rice variety dular of ecotype aus. *Yi Chuan Xue Bao* 27, 409-417.

Yan, C.J., Liang, G.H., Chen, F., Li, X., Tang, S.Z., Yi, C.D., Tan, S., Lu, J.F., and Gu, M.H. (2003). Mapping quantitative trait loci associated with rice grain shape based on an indica/japonica backcross population. *Yi Chuan Xue Bao* 30, 711-716.

Yan, C.J., Liang, G.H., Gu, S.L., Yi, C.D., Lu, J.F., Li, X., Tang, S.Z., and Gu, M.H. (2003). Molecular marker analysis and genetic basis for sterility of typical indica/japonica hybrids. *Yi Chuan Xue Bao* 30, 267-276.

Yan, J., Zhu, J., He, C., Benmoussa, M., and Wu, P. (1998). Molecular Dissection of Developmental Behavior of Plant Height in Rice (*Oryza sativa* L.). *Genetics* 150, 1257-1265.

Yan, J., Zhu, J., He, C., Benmoussa, M., and Wu, P. (1999). Molecular marker-assisted dissection of genotype x environment interaction for plant type traits in rice (*Oryza sativa* L.). *Crop Sci.* 39, 538-544.

Yang, G., Xing, Y., Li, S., Ding, J., Yue, B., Deng, K., Li, Y., and Zhu, Y. (2006). Molecular dissection of developmental behavior of tiller number and plant height and their relationship in rice (*Oryza sativa* L.). *Hereditas* 143, 236-245.

Yang, J., Zhu, J., and Williams, R.W. (2007). Mapping the genetic architecture of complex traits in experimental populations. *Bioinformatics* 23, 1527-1536.

Yang, Q.H., Lu, W., Hu, M.L., Wang, C.M., Zhang, R.X., Yano, M., and Wan, J.M. (2003). QTL and epistatic interaction underlying leaf chlorophyll and H2O2 content variation in rice (*Oryza sativa* L.). *Yi Chuan Xue Bao* 30, 245-250.

Yoo, S.C., Cho, S.H., Zhang, H., Paik, H.C., Lee, C.H., Lu, J., Yoo, J.H., Lee, B.W., Koh, H.J., Seo, H.S., and Paek, N.C. (2007). Quantitative trait loci associated with functional stay-green *SNU-SGI* in rice. *Mol. Cells* 24, 83-94.

Yoo, U., Q., Yue, B., Xue, W.Y., Luo, L.J., and Xiong, L.Z. (2006). Identification of quantitative trait loci for ABA sensitivity at seed germination and seedling stages in rice. *Yi Chuan Xue Bao* 33, 532-541.

Yoshida, S., Ikegami, M., Kuzo, J., Sawada, K., Hashimoto, Z., Ishii, T., Nakamura, C., and Kanmijima, O. (2002). QTL Analysis for Plant and Grain Characters of Sake-Breeding Rice Using a Doubled Haploid Population. *Breeding Science* 52, 309-317.

Yu, B., Lin, Z., Li, H., Li, X., Li, J., Wang, Y., Zhang, X., Zhu, Z., Zhai, W., Wang, X., Xie, D., and Sun, C. (2007). TAC1, a major quantitative trait locus controlling tiller angle in rice. *Plant J.* 52, 891-898.

Yu, S.B., Li, J.X., Xu, C.G., Tan, Y.F., Gao, Y.J., Li, X.H., and Zhang, Q. (1997). Importance of epistasis as the genetic basis of heterosis in an elite rice hybrid. *PNAS* 94, 9236-9231.

Yu, C.Y., Wei, J., Chen, L.M., Jiang, L., Zhai, H.Q., and Wan, J.M. (2002). Identification of quantitative trait loci and epistatic interactions for plant height and heading date in rice. *Theor. Appl. Genet.* 104, 619-625.

Yu, C.Y., Wei, J., Chen, L.M., Jiang, L., Zhai, H.Q., and Wan, J.M. (2005). Identification of a dominant suppressor of photoperiod-sensitive gene using indica/japonica backcrossed progenies in rice (*Oryza sativa* L.). *Rice Genetics Newsletter* 22, 54-.

Zhang, Z.-H., Qu, X.-S., Wan, S., Chen, L.-H., and Zhu, Y.-G. (2005). Comparison of QTL Controlling Seeding Vigour under Different Temperature Conditions Using Recombinant Inbred Lines in Rice (*Oryza sativa*). *Ann. Bot.* 95, 423-429.

Zheng, B.S., Yang, L., Mao, C.Z., Zhang, W.P., and Wu, P. (2006). QTLs and candidate genes for rice root growth under flooding and upland conditions. *Yi Chuan Xue Bao* 33, 141-151.

Zheng, H.G., Babu, R.C., Pathan, H.S., Ali, L., Huang, N., Courtis, B., and Nguyen, H.T. (2000). Quantitative trait loci for root-penetration ability and root thickness in rice: comparison of genetic backgrounds. *Genome* 43, 53-61.

Zhuang, J.Y., Fan, Y.Y., Rao, Z.M., Wu, J.L., Xia, Y.W., and Zheng, K.L. (2002). Analysis on additive effects and additive-by-additive epistatic effects of QTLs for yield traits in a recombinant inbred line population of rice. *Theor. Appl. Genet.* 105, 1137-1145.

Zhuang, J.Y., Fan, Y.Y., Wu, J.L., Xia, Y.W., and Zheng, K.L. (2000). Comparison of QTL detection for yield traits using F2 and recombinant inbred lines in rice. *Rice Genetics Newsletter* 17, 49-51.

Zhuang, J.Y., Fan, Y.Y., Wu, J.L., Xia, Y.W., and Zheng, K.L. (2001). Comparison of the detection of QTL for yield traits in different generations of a rice cross using two mapping approaches. *Yi Chuan Xue Bao* 28, 458-464.

Zhuang, J.Y., Lin, H.X., Lu, J., Qian, H.R., Hittalmani, S., Huang, N., and Zheng, K.L. (1996). Analysis of QTL x environment interaction for yield components and plant height in rice. *Rice Genetics Newsletter* 13, 127-129.

Zhuang, J.Y., Wu, J.-L., Fan, Y.-Y., Rao, Z.-M., and Zheng, K.-L. (2001). Genetic drag between a blast resistance gene and QTL conditioning yield trait detected in a recombinant inbred line population in rice. *Rice Genetics Newsletter* 18, 69-70.

Suppl Table S5. Percent additive effect of each yield-related traits to its parent haplotype (AiAi) and yield traits in BCF1 of MT-RILs.

Populatio	QTL	Marker	Haplotype		n	PN		SN		SF		TGW		GYP		PYP		
						A (%)	P (T<t)	A (%)	P (T<t)	A (%)	P (T<t)	A (%)	P (T<t)	A (%)	P (T<t)	A (%)	P (T<t)	
MT	<i>qGYP1.2</i>	RM259	AiAi	RM600	AiAj	15	1.41	0.331	6.36	0.142	1.48	0.447	-2.18	0.111	7.35	0.197	6.39	0.132
			AiAj	AiAi	10	-0.79	0.438	-1.56	0.399	15.51	0.121	-1.03	0.325	15.73	0.062	-0.62	0.456	
			AiAj	AiAj	64	7.04	0.003	-12.10	0.000	19.66	0.001	0.39	0.354	10.14	0.015	-7.47	0.023	
	<i>qGYP5.1</i>	S05095	AiAi	RM480	AiAj	20	-1.96	0.278	4.03	0.177	0.29	0.488	-2.94	0.024	0.52	0.469	-0.80	0.442
			AiAj	AiAi	21	-0.08	0.490	2.68	0.309	1.06	0.451	-3.87	0.003	1.61	0.401	-0.84	0.439	
			AiAj	AiAj	48	3.62	0.078	2.28	0.260	10.38	0.051	-3.18	0.001	12.70	0.004	-0.98	0.396	
	<i>qGYP6.1</i>	S06015	AiAi	S06018	AiAj	1	-1.99	n.a.	7.61	n.a.	1.37	n.a.	4.00	n.a.	4.26	n.a.	2.09	n.a.
			AiAj	AiAi	7	3.84	0.185	14.82	0.081	-25.43	0.000	3.77	0.052	-5.79	0.150	26.13	0.006	
			AiAj	AiAj	103	2.41	0.165	13.51	0.000	-36.74	0.000	1.22	0.126	-18.46	0.000	30.69	0.000	
	<i>qPYP6.1</i>	RM50	AiAi	RM527	AiAj	9	4.00	0.207	20.59	0.005	-27.18	0.009	-0.92	0.275	-8.79	0.198	26.69	0.003
			AiAj	AiAi	23	-2.19	0.240	16.64	0.000	-33.69	0.000	1.48	0.156	-16.45	0.000	24.27	0.000	
			AiAj	AiAj	53	-2.50	0.133	19.35	0.000	-35.15	0.000	-0.76	0.228	-13.81	0.000	33.29	0.000	
	<i>qGYP8.1</i>	S08059	AiAi	S08066	AiAj	5	3.95	0.176	-7.58	0.149	-17.53	0.135	0.47	0.170	-19.10	0.290	2.27	0.435
			AiAj	AiAi	3	6.62	0.253	-4.98	0.229	29.88	0.091	-3.74	0.286	25.63	0.164	-8.21	0.202	
			AiAj	AiAj	75	-0.83	0.352	-7.10	0.010	12.02	0.019	-0.16	0.433	3.58	0.191	-8.97	0.003	
	<i>qGYP10.1</i>	S10019	AiAi	S10026C	AiAj	5	-12.34	0.048	13.36	0.143	-4.86	0.423	-4.59	0.130	-7.36	0.323	1.73	0.437
			AiAj	AiAi	12	-2.52	0.295	-1.40	0.380	-2.90	0.388	-2.44	0.113	-10.85	0.109	-10.79	0.017	
			AiAj	AiAj	53	0.93	0.338	-4.44	0.085	0.49	0.467	0.51	0.310	-6.06	0.069	-6.11	0.043	
	<i>qPYP11.1</i>	S11112	AiAi	S11117B	AiAj	3	-4.33	0.291	-1.85	0.446	-32.73	0.000	-0.36	0.406	-23.57	0.065	9.27	0.269
			AiAj	AiAi	3	-6.31	0.091	1.52	0.412	-4.00	0.441	-4.41	0.110	-14.29	0.209	-9.13	0.223	
			AiAj	AiAj	61	-2.03	0.173	0.15	0.481	-11.72	0.015	-0.17	0.427	-6.79	0.049	7.26	0.025	

^a percent additive effect of the corresponding haplotype to its parent haplotype (AiAi)^b probability of significance assuming $H_0=0$ by Student's t-test.

Suppl Table S6. Standardized direct effect of yield-related traits to yield in MT-RILs.

QTL	Allele type	DTH	PN	SN	SF	TGW	Most positively effective trait (>0.5 or <-0.5)
<i>qGYP1.2</i>	AiAi/AiAi	-0.027	0.268	0.473	0.897	0.157	SF
	AiAi/AiAj	0.029	-0.104	0.237	1.073	0.233	SF
	AiAj/AiAi	0.388	0.145	0.051	0.897	0.093	SF
	AiAj/AiAj	-0.029	0.263	0.541	0.936	0.185	SN/SF
<i>qGYP5.1</i>	AiAi/AiAi	0.023	0.192	0.359	0.98	0.073	SF
	AiAi/AiAj	-0.03	0.192	0.428	0.911	0.199	SF
	AiAj/AiAi	0.347	0.475	0.335	1.077	0.083	SF
	AiAj/AiAj	0.02	0.254	0.495	0.953	0.146	SF
<i>qPYP6.1</i>	AiAi/AiAi	-0.005	0.271	0.665	-0.336	0.291	SN
	AiAi/AiAj	0.679	0.698	-0.087	-0.515	1.027	DTH/PN/TGW, (SF)
	AiAj/AiAi	0.191	0.746	0.469	-0.288	0.292	PN
	AiAj/AiAj	-0.157	0.34	0.58	-0.467	0.086	SN
<i>qGYP8.1</i>	AiAi/AiAi	0.019	0.303	0.394	0.968	0.204	SF
	AiAi/AiAj	n.a.	n.a.	n.a.	n.a.	n.a.	
	AiAj/AiAi	n.a.	n.a.	n.a.	n.a.	n.a.	
	AiAj/AiAj	0.043	0.208	0.424	0.933	0.021	SF
<i>qPYP11.1</i>	AiAi/AiAi	-0.024	0.292	0.595	-0.408	0.137	SN
	AiAi/AiAj	n.a.	n.a.	n.a.	n.a.	n.a.	
	AiAj/AiAi	n.a.	n.a.	n.a.	n.a.	n.a.	
	AiAj/AiAj	0.154	0.271	0.445	-0.478	0.165	
<i>qGYP6.1</i>	AiAi/AiAi	0.044	0.175	0.424	0.907	0.049	SF
	AiAi/AiAj	n.a.	n.a.	n.a.	n.a.	n.a.	
	AiAj/AiAi	-0.014	-0.562	0.813	-0.572	1.475	SN/TGW, (SF)
	AiAj/AiAj	0.102	0.289	0.41	0.813	0.162	SF
<i>qGYP10.1</i>	AiAi/AiAi	0.183	0.338	0.355	0.951	0.115	SF
	AiAi/AiAj	n.a.	n.a.	n.a.	n.a.	n.a.	
	AiAj/AiAi	0.203	0.449	0.485	1.094	0.35	SF
	AiAj/AiAj	-0.162	0.137	0.545	0.984	0.159	SN/SF

^a effective trait below -0.5 in parenthesis